1125



Intact protein analysis

FEE SCHEDULE 2024 | PROTEOMICS CENTER

ERASMUS MC - ROTTERDAM

		EUR
rotein digestion, sample cleanup		
	o 1-5 gel slice samples	115 / sample
	o 6-10 gel slice samples	75 / sample
	o >11 gel slice samples	45 / sample
	o complete SDS-PAGE lane	450
	o other proteases (V8, Lys-N, Lys-C, pepsin, etc.)	225
	O In-solution digestion, 1-5 samples	115
	o In-solution digestion, >6 samples	75
	On-bead digestion	115
Peptide fractionation of complex mixtures (WCE, issue, etc.)		
•	o HILIC	375
	○ RP, high pH elution	450
	o SCX, SAX	450
	o enrichment for phosphoproteomics (TiO ₂ , IMAC, etc.) o diGly, acetylated, methylated, etc. peptide enrichment	375
		375
	with PTMscan (including reagents), per IP	825
	o diGly, acetylated, methylated, peptide enrichment with PTMscan (excluding reagents)	375
	o other PTM enrichment	enquire
Post-digestion labeling for quantitation (WCE, issue, protein complex, etc.)		
	o TMTPro (16- or 18-plex) labeling per sample	enquire
	o dimethyl labeling per sample	enquire
	o ¹⁸ O labeling per sample	enquire
All additional, non-standard sample handling will		
	Mass spectrometry	
LC-MS (DDA/DIA on the Orbitrap Fusion Lumos Tribrid, Orbitrap Eclipse, Orbitrap Exploris 480 or	Mass spectrometry	
Tribrid, Orbitrap Eclipse, Orbitrap Exploris 480 or	Mass spectrometry	
Tribrid, Orbitrap Eclipse, Orbitrap Exploris 480 or	Mass spectrometry 1 LC-MS (RP) run 60 min	135
· · · · · · · · · · · · · · · · · · ·		135 200
Tribrid, Orbitrap Eclipse, Orbitrap Exploris 480 or	1 LC-MS (RP) run 60 min	
Fribrid, Orbitrap Eclipse, Orbitrap Exploris 480 or	1 LC-MS (RP) run 60 min 1 LC-MS (RP) run 120 min	200

Data analysis

Standard data analysis includes identification and quantitation (SILAC, TMT and LFQ) for DDA and DIA analyses. The choice of software tool depends on the experimental design. We use MaxQuant, Proteome Discoverer / Mascot, PEAKS, Spectronaut, DIA-NN, Byonic, etc.

Additional downstream data analyses (statistical analysis, Perseus, etc.)	95 / hour

Two example applications:

Example 1: Complete gel lane

10x LC-MS run (60 min)	1350
Total	1800

Example 2: In-solution digest + HILIC fractionation of a complex sample

Total	1690
6x LC-MS run (120 min)	1200
HILIC fractionation	375
Protein digestion	115

Terms & conditions

- Consultation regarding study design, data interpretation, and publication / public access is provided at no cost. Please note that charges include only a *minor* part of the reagent and personnel costs – the major part is covered by Erasmus MC core funding. The Proteomics Center realizes no profit whatsoever.
- The Proteomics Center runs all projects on a collaborative basis, which is expected to result in coauthorship if data generated by the Center are published in scientific papers. This scientific output allows our scientists to apply for research and infrastructure funding, which will ensure access to the state-of-the-art proteomics technology for Erasmus MC and external researchers.
- New collaborators are allowed to run one pilot experiment for free. If the results are satisfactory for both parties, the costs for subsequent experiments as indicated above will be charged to the collaborator.

For further information please contact the core facility director, Dr. Jeroen Demmers | email: j.demmers@erasmusmc.nl | https://proteomics.center

Visiting address
Proteomics Center
Erasmus University Medical Center
Room Ee-679A | Faculty Building Ee, 6th Floor
Wytemaweg 80
3015 CN Rotterdam
the Netherlands